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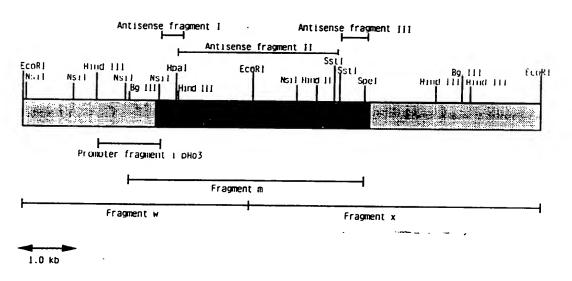
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(54) Title: GENETICALLY ENGINEERED MODIFICATION OF POTATO TO FORM AMYLOPECTIN-TYPE STARCH

Result of restriction analysis. GBSS coding region including introns are marked in a darker tone.



(57) Abstract

Genetically engineered modification of potato for suppressing the formation of amylose-type starch is described. Three fragments for insertion in the antisense direction into the potato genome are also described. Moreover, antisense constructs, genes and vectors comprising said antisense fragments are described. Further a promoter for the gene coding for formation of granulebound starch synthase and also the gene itself are described. Also cells, plants, tubers, microtubers and seeds of potato comprising said antisense fragments are described. Finally, amylopectin-type starch, both native and derivatised, derived from the potato that is modified in a genetically engineered manner, as well as a method of suppressing amylose formation in potato are described.



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GENETICALLY ENGINEERED MODIFICATION OF POTATO TO FORM AMYLOPECTIN-TYPE STARCH

The present invention relates to genetically engi-5 neered modification of potato, resulting in the formation of practically solely amylopectin-type starch in the potato. The genetically engineered modification implies the insertion of gene fragments into potato, said gene fragments comprising parts of leader sequence, translation 10 start, translation end and trailer sequence as well as coding and noncoding (i.e. exons and introns) parts of the gene for granule-bound starch synthase, inserted in the antisense direction.

Background of the Invention

Starch in various forms is of great import in the food and paper industry. In future, starch will also be a great potential for producing polymers which are degradable in nature, e.g. for use as packing material. Many different starch products are known which are produced by 20 derivatisation of native starch originating from, inter alia, maize and potato. Starch from potato and maize, respectively, is competing in most market areas.

In the potato tuber, starch is the greatest part of the solid matter. About 1/4 to 1/5 of the starch in potato is amylose, while the remainder of the starch is amylopectin. These two components of the starch have different fields of application, and therefore the possibility of producing either pure amylose or pure amylopectin is most interesting. The two starch components can be produced from common starch, which requires a number of process steps and, consequently, is expensive and complicated.

It has now proved that by genetic engineering it is possible to modify potato so that the tubers merely produce mainly starch of one or the other type. As a result, 35 a starch quality is obtained which can compete in the areas where potato starch is normally not used today. Starch from such potato which is modified in a genetically

engineered manner has great potential as a food additive, since it has not been subjected to any chemical modification process.

Starch Synthesis

5 The synthesis of starch and the regulation thereof are presently being studied with great interest, both on the level of basic research and for industrial application. Although much is known about the assistance of certain enzymes in the transformation of saccharose into starch, the biosynthesis of starch has not yet been elucidated. By making researches above all into maize, it has, however, been possible to elucidate part of the ways of synthesis and the enzymes participating in these reactions. The most important starch-synthesising enzymes for 15 producing the starch granules are the starch synthase and the branching enzyme. In maize, three forms of starch synthase have so far been demonstrated and studied, two of which are soluble and one is insolubly associated with the starch granules. Also the branching enzyme consists of 20 three forms which are probably coded by three different genes (Mac Donald & Preiss, 1985; Preiss, 1988).

The Waxy Gene in Maize

The synthesis of the starch component amylose essentially occurs by the action of the starch synthase alpha-1,4-D-glucane-4-alpha-glucosyl transferase (EC 2.4.1.21) which is associated with the starch granules in the growth cell. The gene coding for this granule-bound enzyme is called "waxy" (= \underline{wx}^+), while the enzyme is called "GBSS" (granule-bound starch synthase).

waxy locus in maize has been thoroughly characterised both genetically and biochemically. The waxy gene on chromosome 9 controls the production of amylose in endosperm, pollen and the embryo sac. The starch formed in endosperm in normal maize with the wx⁺ allele consists to 25% of amylose and to 75% of amylopectin. A mutant form of maize has been found in which the endosperm contains a mutation located to the wx⁺ gene, and therefore no functioning GBSS

is synthesised. Endosperm from this mutant maize therefore contains merely amylopectin as the starch component. This so-called waxy mutant thus contains neither GBSS nor amylose (Echt & Schwartz, 1981).

The GBSS protein is coded by the wx gene in the cell nucleus but is transported to and active in the amyloplast. The preprotein therefore consists of two components, viz. a 7 kD transit peptide which transfers the protein across the amyloplast membrane, and the actual 10 protein which is 58 kD. The coding region of the wx gene in maize is 3.7 kb long and comprises 14 exons and 13 introns. A number of the regulation signals in the promoter region are known, and two different polyadenylating sequences have been described (Klösgen et al, 1986; 15 Schwartz-Sommer et al, 1984; Shure et al, 1983).

Amylose Enzyme in Potato

In potato, a 60 kD protein has been identified, which constitutes the main granule-bound protein. Since antibodies against this potato enzyme cross-react with GBSS from 20 maize, it is assumed that it is the granule-bound synthase (Vos-Scheperkeuter et al, 1986). The gene for potato GBSS has, however, so far not been characterised to the same extent as the waxy gene in maize, either in respect of locating or structure.

25 Naturally occurring waxy mutants have been described for barley, rice and sorghum besides maize. In potato no natural mutant has been found, but a mutant has been produced by X-radiation of leaves from a monohaploid (n=12) plant (Visser et al, 1987). Starch isolated from tubers of this mutant contains neither the GBSS protein nor amylose. The mutant is conditioned by a simple recessive gene and is called amf. It may be compared to waxy mutants of other plant species since both the GBSS protein and amylose are lacking. The stability of the chromosome number, however, 35 is weakened since this is quadrupled to the natural number (n=48), which can give negative effects on the potato plants (Jacobsen et al, 1990).

Inhibition of Amylose Production

The synthesis of amylose can be drastically reduced by inhibition of the granule-bound starch synthase, GBSS, which catalyses the formation of amylose. This inhibition results in the starch mainly being amylopectin.

Inhibition of the formation of enzyme can be accomplished in several ways, e.g. by:

- mutagen treatment which results in a modification of the gene sequence coding for the formation of the enzyme
- 10 incorporation of a transposon in the gene sequence coding for the enzyme
 - genetically engineered modification so that the gene coding for the enzyme is not expressed, e.g. antisense gene inhibition.
- Fig. 1 illustrates a specific suppression of normal gene expression in that a complementary antisense nucleotide is allowed to hybridise with mRNA for a target gene. The antisense nucleotide thus is antisense RNA which is transcribed in vivo from a "reversed" gene sequence 20 (Izant, 1989).

By using the antisense technique, various gene functions in plants have been inhibited. The antisense construct for chalcone synthase, polygalacturonase and phosphinotricin acetyltransferase has been used to inhibit the corresponding enzyme in the plant species petunia, tomato and tobacco.

Inhibition of Amylose in Potato

In potato, experiments have previously been made to inhibit the synthesis of the granule-bound starch synthase (GBSS protein) with an antisense construct corresponding to the gene coding for GBSS (this gene is hereinafter called the "GBSS gene"). Hergersberger (1988) describes a method by which a cDNA clone for the GBSS gene in potato has been isolated by means of a cDNA clone for the wx⁺ gene in maiz . An antisense construct based on the entire cDNA clone was transferred to leaf discs of potato by means of Agrobacterium tumefaciens. In microtubers induced

in vitro from regenerated potato sprouts, a varying and very weak reduction of the amylose content was observed and shown in a diagram. A complete characterisation of the GBSS gene is not provided.

The gene for the GBSS protein in potato has been further characterised in that a genomic wx⁺ clone was examined by restriction analysis. However, the DNA sequence of the clone has not been determined (Visser et al, 1989).

Further experiments with an antisense construct corresponding to the GBSS gene in potato have been reported.
The antisense construct which is based on a cDNA clone
together with the CaMV 35S promoter has been transformed
by means of Agrobacterium rhizogenes. According to information, the transformation resulted in a lower amylose
content in the potato, but no values have been accounted
for (Flavell, 1990).

None of the methods used so far for genetically engineered modification of potato has resulted in potato with practically no amylose-type starch.

The object of the invention therefore is to provide a practically complete suppression of the formation of amylose in potato tubers.

Summary of the Invention

According to the invention, the function of the GBSS 25 gene and, thus, the amylose production in potato are inhibited by using completely new antisense constructs. For forming the antisense fragments according to the invention, the genomic GBSS gene is used as a basis in order to achieve an inhibition of GBSS and, consequently, of the 30 amylose production, which is as effective as possible. The antisense constructs according to the invention comprise both coding and noncoding parts of the GBSS gene which correspond to sequences in the region comprising promoter as well as leader sequence, translation start, translation 35 end and trailer sequence in the antisense direction. For a tissue-specific expression, i.e. the amylose production should be inhibited in the potato tubers only, use is made

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of promoters which are specifically active in the potato tuber. As a result, the starch composition in other parts of the plant is not affected, which otherwise would give negative side-effects.

The invention thus comprises a fragment which essentially has one of the nucleotide sequences stated in SEQ ID No. 1, SEQ ID No. 2 or SEQ ID No. 3. However, the sequences may deviate from those stated by one or more non-adjacent base pairs, without affecting the function of the fragments.

The invention also comprises a potato-tuber-specific promoter comprising 987 bp which belongs to the gene according to the invention, which codes for granule-bound starch synthase. Neither the promoter nor the corresponding gene has previously been characterised. The promoter sequence of 987 bp is stated in SEQ ID No. 4, while the gene sequence is stated in SEQ ID No. 5. Also the promoter and gene sequences may deviate from those stated by one or more non-adjacent base pairs, without affecting their function.

The invention also comprises vectors including the antisense fragments and the antisense constructs according to the invention.

In other aspects the invention comprises cells,
25 plants, tubers, microtubers and seeds whose genome contains the fragments according to the invention inserted in the antisense direction.

In still further aspects, the invention comprises amylopectin-type starch, both native and derivatised.

Finally, the invention comprises a method of suppressing amylose formation in potato, whereby mainly amylopectin-type starch is formed in the potato.

The invention will now be described in more detail with reference to the accompanying Figures in which

Fig. 1 illustrates the principle of the antisense gene inhibition,

Fig. 2 shows the result of restriction analysis of the potato GBSS gene,

Fig. 3 shows two new binary vectors pHo3 and pHo4,

Fig. 4 shows the antisense constructs pHoxwA, pHoxwB and pHoxwD,

Fig. 5 shows the antisense constructs pHoxwF and pHoxwG, and

Fig. 6 shows the antisense constructs pHoxwK and pHoxwL.

Moreover, the sequences of the different DNA fragments according to the invention are shown in SEQ ID Nos 1, 2, 3, 4 and 5. There may be deviations from these sequences in one or more non-adjacent base pairs. MATERIALS

In the practical carrying out of the invention the following materials were used:

Bacterial strains: E. coli DH5alfa and DH5alfaF'IQ(BRL). E. coli JM105 (Pharmacia). A. tumefaciens LBA4404 (Clontech).

20 <u>Vectors</u>: M13mp18 and mp19 (Pharmacia). pBI101 and pBI121 (Clontech). pBI240.7 (M. W. Bevan). pUC plasmids (Pharmacia).

Enzymes: Restriction enzymes and EcoRI linker (BRL). UNION TM DNA Ligation Kit (Clontech). Sequenase TM DNA

25 Sequencing Kit (USB). T₄-DNA ligase (Pharmacia).

The above-mentioned materials are used according to specifications stated by the manufacturers.

Genomic Library

A genomic library in EMBL3 has been produced by Clon-30 tech on the applicant's account, while using leaves of the potato Bintje as starting material.

Identification and Isolation of the GBSS Gene

The genomic library has been screened for the potato GBSS gene by means of cDNA clones for both the 5' and 3' end of the gene (said cDNA clones being obtained from M Hergersberger, Max Plank Institute in Cologne) according to a protocol from Clontech.

A full-length clone of the potato GBSS gene, wx311, has been identified and isolated from the genomic library. The start of the GBSS gene has been determined at an EcoRI fragment which is called fragment w (3.95 kb). The end of the GBSS gene has also been determined at an EcoRI fragment which is called fragment x (5.0 kb). A BgIII-SpeI fragment which is called fragment m (3.9 kb) has also been isolated and shares sequences both from fragment w and from fragment x. The fragments w, m and x have been sub-cloned in pUC13 (Viera, 1982; Yanisch-Peron et al, 1985) and are called pSw, pSm and pSx, respectively (Fig. 2). Characterisation of the GBSS Gene in Potato

The GBSS gene in potato has been characterised by restriction analysis and cDNA probes, where the 5' and 3' end of the GBSS gene has been determined more accurately (Fig. 2). Sequence determination according to Sanger et al, 1977 of the GBSS gene has been made on subclones from pSw and pSx in M13mp18 and mp19 as well as pUC19 starting around the 5' end (see SEQ ID No. 5).

20 The promoter region has been determined at a BglII-NsiI fragment (see SEQ ID No. 4). Transcription and translation start has been determined at an overlapping BglII-HindIII fragment. The terminator region has in turn been determined at a SpeI-HindIII fragment.

25 Antisense Constructs for the GBSS Gene in Potato

The GBSS gene fragments according to the invention (see SEQ ID Nos 1, 2 and 3, and Fig. 2) have been determined in the following manner.

The restriction of pSw with NsiI and HindIII gives

fragment I (SEQ ID No. 1) which subcloned in pUC19 is
called 19NH35. Further restriction of 19 NH35 with HpaISstI gives a fragment containing 342 bp of the GBSS gene
according to the invention. This fragment comprises leader
sequence, translation start and the first 125 bp of the

coding region.

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The restriction of pSm with HpaI and NsiI gives fragment II (SEQ ID No. 2) which subcloned in pJRD184 (Heusterspreute et al, 1987) is called pJRDmitt. Further restriction of pJRDmitt with HpaI-SstI gives a fragment 5 containing 2549 bp of the GBSS gene according to the invention. This fragment comprises exons and introns from the middle of the gene.

The restriction of pSx with SstI and SpeI gives fragment III (SEQ ID No. 3) which subcloned in pBluescript

10 (Melton et al, 1984) is called pBlue3'. Further restriction of pBlue3' with BamHI-SstI gives a fragment containing 492 bp of the GESS gene according to the invention.

This fragment comprises the last intron and exon, translation end and 278 bp of trailer sequence.

Antisense Constructs with Fragment I (Fig. 4): For the antisense construct pHoxwA, the HpaI-SstI fragment from 19NH35 has been inserted in the antisense direction into the binary vector pBI121 (Jefferson et al, 1987) cleaved with SmaI-SstI. The transcription of the antisense fragment is then initiated by the CaMV 35S promoter and is terminated by the NOS terminator (NOS = nopaline synthase).

For the antisense construct pHoxwB, the HpaI-SstI fragment from 19NH35 has been inserted in the antisense

25 direction into the binary vector pHo4 (Fig. 3) cleaved with SmaI-SstI. The patatin I promoter which is tuber specific in potato comes from the vector pBI240.7 obtained from M. Bevan, Institute of Plant Science Research, Norwich. The transcription of the antisense fragment is then initiated by the patatin I promoter and is terminated by the NOS terminator.

For the antisense construct pHoxwD, the HpaI-SstI fragment from 19NH35 has been inserted in the antisense direction into the binary vector pHo3 (Fig. 3) cleaved

35 with SmaI-SstI. pHo3 is a new binary vector which is constructed on the basis of pBI101. This vector which contains the promoter according to the invention (see SEQ ID

No. 4) (GBSS promoter) of the now characterised potato GBSS gene according to the invention has been restriction-cleaved with SmaI and SstI, the HpaI-SstI fragment from 19NH35 being inserted in the antisense direction. The transcription of the antisense fragment is then initiated by its own GBSS promoter and is terminated by the NOS terminator. This means that the antisense fragment is transcribed only in the potato tuber, since the GBSS promoter like the patatin I promoter is tuber-specific.

Antisense Constructs with Fragment II (Fig. 5): For the antisense construct pHoxwF, the HpaI-SstI fragment from pJRDmitt has been inserted in the antisense direction into the binary vector pHo4 cleaved with SmaI-SstI. The transcription of the antisense fragment is then initiated by the patatin I promoter and terminated by the NOS terminator.

For the antisense construct pHoxwG, the HpaI-SstI fragment from pJRDmitt has been inserted in the antisense direction into the binary vector pHo3 cleaved with SmaI-SstI. The transcription of the antisense fragment is then initiated by its own GBSS promoter and is terminated by the NOS terminator.

Antisense Constructs with Fragment III (Fig. 6): For the antisense construct pHoxwK, the BamHI-SstI fragment from pBlue3' has been inserted in the antisense direction into the binary vector pHo4 cleaved with BamHI-SstI. The transcription of the antisense fragment is then initiated by the patatin I promoter and is terminated by the NOS terminator.

fragment from pBlue3' has been inserted in the antisense direction into the binary vector pHo3 cleaved with BamHI-SstI. The transcription of the antisense fragment is then initiated by its own GBSS promoter and is terminated by the NOS terminator.

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The formed antisense constructs (Figs 4, 5, 6) have been transformed to Agrobacterium tumefaciens strain LBA4404 by direct transformation with the "freeze-thawing" method (Hoekema et al, 1983; An et al, 1988).

5 Transformation

The antisense constructs are transferred to bacteria, suitably by the "freeze-thawing" method (An et al, 1988). The transfer of the recombinant bacterium to potato tissue occurs by incubation of the potato tissue with the recom-10 binant bacterium in a suitable medium after some sort of damage has been inflicted upon the potato tissue. During the incubation, T-DNA from the bacterium enters the DNA of the host plant. After the incubation, the bacteria are killed and the potato tissue is transferred to a solid 15 medium for callus induction and is incubated for growth of callus.

After passing through further suitable media, sprouts are formed which are cut away from the potato tissue.

Checks for testing the expression of the antisense 20 constructs and the transfer thereof to the potato genome are carried out by e.g. southern and northern hybridisation (Maniatis et al (1982)). The number of copies of the antisense construct which has been transferred is determined by southern hybridisation.

The testing of the expression on protein level is suitably carried out on microtubers induced in vitro on the transformed sprouts, thus permitting the testing to be performed as quickly as possible.

Characterisation of the GBSS Protein

The effect of the antisense constructs on the function of the GBSS gene with respect to the activity of the GBSS protein is examined by extracting starch from the microtubers and analysing it regarding the presence of the GBSS protein. In electrophoresis on polyacrylamide gel 35 (Hovenkamp-Hermelink et al, 1987), the GBSS protein forms a distinct band at 60 kD, when the GBSS gene functions. When the GBSS gene is not expressed, i.e. when the anti-

sense GBSS gene is fully expressed, thereby inhibiting the formation of GBSS protein, no 60 kD band is demonstrated on the gel.

Characterisation of the Starch

The composition of the starch in microtubers is identical with that of ordinary potato tubers, and therefore the effect of the antisense constructs on the amylose production is examined in microtubers. The proportion of amylose to amylopectin can be determined by a spectrophoto-10 metric method (e.g. according to Hovenkamp-Hermelink et al, 1988).

Extraction of Amylopectin from Amylopectin Potato

Amylopectin is extracted from the so-called amylopectin potato (potato in which the formation of amylose 15 has been suppressed by inserting the antisense constructs according to the invention) in a known manner.

Derivatisation of Amylopectin

Depending on the final use of the amylopectin, its physical and chemical qualities can be modified by deri-20 vatisation. By derivatisation is here meant chemical, physical and enzymatic treatment and combinations thereof (modified starches).

The chemical derivatisation, i.e. chemical modification of the amylopectin, can be carried out in different 25 ways, for example by oxidation, acid hydrolysis, dextrinisation, different forms of etherification, such as cationisation, hydroxy propylation and hydroxy ethylation, different forms of esterification, for example by vinyl acetate, acetic anhydride, or by monophosphatising, 30 diphosphatising and octenyl succination, and combinations thereof.

Physical modification of the amylopectin can be effected by e.g. cylinder-drying or extrusion.

In enzymatic derivatisation, degradation (reduction 35 of the viscosity) and chemical modification of the amylopectin are effected by means of existing enzymatic systems.

Th derivatisation is effected at different temperatures, according to the desired end product. The ordinary range of temperature which is used is $20-45\,^{\circ}\text{C}$, but temperatures up to $180\,^{\circ}\text{C}$ are possible.

5 The invention will be described in more detail in the following Examples.

Example 1

Production of microtubers with inserted antisense constructs according to the invention

The antisense constructs (see Figs 4, 5 and 6) are transferred to Agrobacterium tumefaciens LBA 4404 by the "freeze-thawing" method (An et al, 1988). The transfer to potato tissue is carried out according to a modified protocol from Rocha-Sosa et al (1989).

Leaf discs from potato plants cultured in vitro are incubated in darkness on a liquid MS-medium (Murashige & Skoog; 1962) with 3% saccharose and 0.5% MES together with 100 µl of a suspension of recombinant Agrobacterium per 10 ml medium for two days. After these two days the bacteria are killed. The leaf discs are transferred to a solid medium for callus induction and incubated for 4-6 weeks, depending on the growth of callus. The solid medium is composed as follows:

MS + 3% saccarose

25 2 mg/l zeatin riboside

0.02 mg/l "NAA"

0.02 mg/l "GA₂"

500 mg/l "Claforan"

50 mg/l kanamycin

30 0.25% "Gellan"

Subsequently the leaf discs are transferred to a medium having a different composition of hormones, comprising:

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MS + 3% saccharose

5 mg/l "NAA"

0.1 mg/l "BAP"

500 mg/l "Claforan"

50 mg/l kanamycin

0.25% "Gellan"

The leaf discs are stored on this medium for about 4 weeks, whereupon they are transferred to a medium in which the "Claforan" concentration has been reduced to 250 mg/l. If required, the leaf discs are then moved to a fresh medium every 4 or 5 weeks. After the formation of sprouts, these are cut away from the leaf discs and transferred to an identical medium.

The condition that the antisense construct has been transferred to the leaf discs is first checked by analys-15 ing leaf extracts from the regenerated sprouts in respect of glucuronidase activity by means of the substrates described by Jefferson et al (1987). The activity is demonstrated by visual assessment.

Further tests of the expression of the antisense constructs and the transfer thereof to the potato genome are carried out by southern and northern hybridisation according to Maniatis et al (1981). The number of copies of the antisense constructs that has been transferred is deter-25 mined by southern hybridisation.

When it has been established that the antisense constructs have been transferred to and expressed in the potato genome, the testing of the expression on protein level begins. The testing is carried out on microtubers 30 which have been induced in vitro on the transformed sprouts, thereby avoiding the necessity of waiting for the development of a complete potato plant with potato tubers.

Stem pieces of the potato sprouts are cut off at the nodes and placed on a modified MS medium. There they form 35 microtubers after 2-3 weeks in incubation in darkness at 19°C (Bourque et al, 1987). The medium is composed as follows:

MS + 6% saccharose

2.5 mg/l kinetin

2.5 mg/l "Gellan"

The effect of the antisense constructs on the function of the GBSS gene in respect of the activity of the
GBSS protein is analysed by means of electrophoresis on
polyacrylamide gel (Hovenkamp-Hermelink et al, 1987).
Starch is extracted from the microtubers and analysed
regarding the presence of the GBSS protein. In a polyacrylamide gel, the GBSS protein forms a distinct band at
60 kD, when the GBSS gene functions. If the GBSS gene is
not expressed, i.e. when the antisense GBSS gene is fully
expressed so that the formation of GBSS protein is inhibited, no 60 kD band can be seen on the gel.

The composition of the starch, i.e. the proportion of amylose to amylopectin, is determined by a spectrophotometric method according to Hovenkamp-Hermelink et al (1988), the content of each starch component being determined on the basis of a standard graph.

20 Example 2

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Extraction of amylopectin from amylopectin potato.

Potato whose main starch component is amylopectin, below called amylopectin potato, modified in a genetically engineered manner according to the invention, is grated, thereby releasing the starch from the cell walls.

The cell walls (fibres) are separated from fruit juice and starch in centrifugal screens (centrisiler). The fruit juice is separated from the starch in two steps, viz. first in hydrocyclones and subsequently in specially designed band-type vacuum filters.

Then a finishing refining is carried out in hydrocyclones in which the remainder of the fruit juice and fibres are separated.

The product is dried in two steps, first by predrying on a vacuum filter and subsequently by final drying in a hot-air current.

Example 3

Chemical derivatisation of amylopectin

Amylopectin is sludged in water to a concentration of 20-50%. The pH is adjusted to 10.0-12.0 and a quatenary ammonium compound is added in such a quantity that the end product obtains a degree of substitution of 0.004-0.2. The reaction temperature is set at 20-45°C. When the reaction is completed, the pH is adjusted to 4-8, whereupon the product is washed and dried. In this manner the cationic starch derivative 2-hydroxy-3-trimethyl ammonium propyl ether is obtained.

Example 4

Chemical derivatisation of amylopectin

Amylopectin is sludged in water to a water content

of 10-25% by weight. The pH is adjusted to 10.0-12.0,
and a quatenary ammonium compound is added in such a quantity that the end product obtains a degree of substitution of 0.004-0.2. The reaction temperature is set at 20-45°C.
When the reaction is completed, the pH is adjusted to 4-8.

The end product is 2-hydroxy-3-trimethyl ammonium propyl ether.

Example 5

Chemical derivatisation of amylopectin

Amylopectin is sludged in water to a concentration of 20-50% by weight. The pH is adjusted to 5.0-12.0, and sodium hypochlorite is added so that the end product obtains the desired viscosity. The reaction temperature is set at 20-45°C. When the reaction is completed, the pH is adjusted to 4-8, whereupon the end product is washed and dried. In this manner, oxidised starch is obtained.

Example 6

Physical derivatisation of amylopectin

Amylopectin is sludged in water to a concentration of 20-50% by weight, whereupon the sludge is applied to 35 a heated cylinder where it is dried to a film.

Example 7

Chemical and physical derivatisation of amylopectin

Amylopectin is treated according to the process
described in one of Examples 3-5 for chemical modification and is then further treated according to Example 6
for physical derivatisation.

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SEQ ID No. 1

Sequenced molecule: genomic DNA

Name: GBSS gene fragment from potato

Length of sequence: 342 bp

TGCATGTTTC CCTACATTCT ATTTAGAATC GTGTTGTGGT GTATAAACGT TGTTTCATAT CTCATCTCAT CTATTCTGAT TTTGATTCTC TTGCCTACTG TAATCGGTGA TAAATGTGAA TGCTTCCTTT CTTCTCAGAA ATCAATTTCT GTTTGTTTT TGTTCATCTG TAGCTTATTC TCTGGTAGAT TCCCCTTTTT GTAGACCACA CATCAC ATG GCA AGC ATC ACA GCT TCA CAC CAC Met Ala Ser Ile Thr Ala Ser His His 1 5	50 100 150 200 243
TTT GTG TCA AGA AGC CAA ACT TCA CTA GAC ACC AAA TCA ACC Phe Val Ser Arg Ser Gln Thr Ser Leu Asp Thr Lys Ser Thr 10 15 20	285
TTG TCA CAG ATA GGA CTC AGG AAC CAT ACT CTG ACT CAC AAT Leu Ser Gln Ile Gly Leu Arg Asn His Thr Leu Thr His Asn 25 30 35	327
GGT TTA AGG GCT GTT Gly Leu Arg Ala Val 40	342

. , . , WO 92/11376 PCT/SE91/00892

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SEQ ID No. 2

Sequenced molecule: genomic DNA

Name: GBSS gene fragment from potato

Length of sequence: 2549 bp

AAC AAG CTT GAT GGG CTC CAA TCA ACA ACT AAT ACT AAG GTA Asn Lys Leu Asp Gly Leu Gln Ser Thr Thr Asn Thr Lys Val 45 50 55	42
ACA CCC AAG ATG GCA TCC AGA ACT GAG ACC AAG AGA CCT GGA Thr Pro Lys Met Ala Ser Arg Thr Glu Thr Lys Arg Pro Gly 60 65 70	84
TGC TCA GCT ACC ATT GTT TGT GGA AAG GGA ATG AAC TTG ATC Cys Ser Ala Thr Ile Val Cys Gly Lys Gly Met Asn Leu Ile 75	126
TTT GTG GGT ACT GAG GTT GGT CCT TGG AGC AAA ACT GGT GGA Phe Val Gly Thr Glu Val Gly Pro Trp Ser Lys Thr Gly Gly 85 90 95	168
CTA GGT GAT GTT CTT GGT GGA CTA CCA CCA GCC CTT GCA Leu Gly Asp Val Leu Gly Gly Leu Pro Pro Ala Leu Ala 100 105 110	207
GTAAGTCITT CTITCATTTG GTTACCTACT CATTCATTAC TTATTTTGTT TAGTTAGTTT CTACTGCATC AGTCTTTTTA TCATTTAG GCC CGC GGA Ala Arg Gly	257 304
CAT CGG GTA ATG ACA ATA TCC CCC CGT TAT GAC CAA TAC AAA His Arg Val Met Thr Ile Ser Pro Arg Tyr Asp Gln Tyr Lys 115 120 125	346
GAT GCT IGG GAT ACT GGC GTT GCG GTT GAG GTACATCTTC Asp Ala Trp Asp Thr Gly Val Ala Val Glu 130 135	386
CTATATIGAT ACGGTACAAT ATTGTTCTCT TACATTTCCT GATTCAAGAA IGTGATCATC TGCAG GTC AAA GTT GGA GAC AGC ATT GAA ATT GTT Val Lys Val Gly Asp Ser Ile Glu Ile Val 140 145	436 481
CGT TTO TTT CAC TGC TAT AAA CGT GGG GTT GAT CGT GTT TTT Arg Phe Phe His Cys Tyr Lys Arg Gly Val Asp Arg Val Phe 151 155 160	523
GTT GAO DAO DOA ATG TTO TTG GAG AAA GTAAGCATAT Val Asp His Pro Met Phe Leu Glu Lys 155	560

TATGATIATG AATCCGTCCT GAGGGATACG CAGAACAGGT CATTITGAGT ATCTTTTAAC TCTACTGGTG CTTTTACTCT TTTAAG GTT TGG GGC AAA Val Trp Gly Lys 175	610 658
ACT GGT TCA AAA ATC TAT GGC CCC AAA GCT GGA CTA GAT TAT Thr Gly Ser Lys Ile Tyr Gly Pro Lys Ala Gly Leu Asp Tyr 180	700
CTG GAC AAT GAA CTT AGG TTC AGC TTG TTG TGT CAA Leu Asp Asn Glu Leu Arg Phe Ser Leu Leu Cys Gln 190 195 200	736
GTAAGTTAGT TACTCTTGAT TTTTATGTGG CATTTTACTC TTTTGTCTTT AATCGTTTTT TTAACCTTGT TTTCTCAG GCA GCC CTA GAG GCA CCT Ala Ala Leu Glu Ala Pro 205	786 832
ARA GTT TTG ART TTG ARC AGT AGC ARC TAC TTC TCA GGA CCA Lys Val Leu Asn Leu Asn Ser Ser Asn Tyr Phe Ser Gly Pro 210 215 220	874
TAT G GTAATTAACA CATCCTAGTT TCAGAAAACT CCTTACTATA Tyr G	918
TCATTGTAGG TAATCATCTT TATTTTGCCT ATTCCTGCAG GA GAG GAT ly Glu Asp 225	966
GTT CTC TTC ATT GCC AAT GAT TGG CAC ACA GCT CTC ATT CCT Val Leu Phe Ile Ala Asn Asp Trp His Thr Ala Leu Ile Pro 230 235	1008
TGC TAC TTG AAG TCA ATG TAC CAG TCC AGA GGA ATC TAC TTG Cys Tyr Leu Lys Ser Met Tyr Gln Ser Arg Gly Ile Tyr Leu 240 245	1050
AAT GCC AAG GTAAAATTTC TTTGTATTCA CTCGATTGCA Asn Ala Lys 255	1089
CGTTACCCTG CAAATCAGTA AGGTTGTATT AATATATGAT AAATTTCACA TTGCCTCCAG GTT GCT TTC TGC ATC CAT AAC ATT GCC TAC CAA Val Ala Phe Cys Ile His Asn Ile Ala Tyr Gln 260 265	1139 1182
GGT CGA TIT TOT TTO TOT GAC TTO CCT CTT CTC AAT CTT CCT Gly Arg Phe Ser Phe Ser Asp Phe Pro Leu Leu Asn Leu Pro 270 275 280	1224
GAT GAA TOO AGG GGT TOT TIT GAT TOO ATT GAT GGG TAT Asp Glu Fhe Arg Gly Ser Phe Asp Phe Ile Asp Gly Tyr 255	1263
GTATTTATED TTERARTORS ACCTOCARCT TTTERAGGTO TTTTERTECT	1313

AGTAAATTGA GTTTTTAAAA TTTTGCAGAT ATGAG AAG CCT GTT AAG Lys Pro Val Lys 295	1360
GGT AGG AAA ATC AAC TGG ATG AAG GCT GGG ATA TTA GAA TCA Gly Arg Lys Ile Asn Trp Met Lys Ala Gly Ile Leu Glu Ser 300 305 310	1402
CAT AGG GTG GTT ACA GTG AGC CCA TAC TAT GCC CAA GAA CTT His Arg Val Val Thr Val Ser Pro Tyr Tyr Ala Gln Glu Leu 315	
GTC TCT GCT GTT GAC AAG GGA GTT GAA TTG GAC AGT GTC CTT Val Ser Ala Val Asp Lys Gly Val Glu Leu Asp Ser Val Leu 330 335	1486
CGT AAG ACT TGC ATA ACT GGG ATT GTG AAT GGC ATG GAT ACA Arg Lys Thr Cys Ile Thr Gly Ile Val. Asn Gly Met Asp Thr 345	1528
CAA GAG TGG AAC CCA GCG ACT GAC AAA TAC ACA GAT GTC AAA Gln Glu Trp Asn Pro Ala Thr Asp Lys Tyr Thr Asp Val Lys 355 360 365	
TAC GAT ATA ACC ACT GTAAGATAAG ATTTTTCCGA CTCCAGTATA Tyr Asp Ile Thr Thr 370	1615
TACTAAATTA TTTTGTATGT TTATGAAATT AAAGAGTTCT TGCTAATCAA AATCTCTATA CAG GTC ATG GAC GCA AAA CCT TTA CTA AAG GAG Val Met Asp Ala Lys Pro Leu Leu Lys Glu 375 380	1665 1708
GCT CTT CAA GCA GCA GTT GGC TTG CCT GTT GAC AAG AAG ATG Ala Leu Gln Ala Ala Val Gly Leu Pro Val Asp Lys Ile 385 390 395	
CCT TTG ATT GGC TTC ATC GGC AGA CTT GAG GAG CAG AAA GGT Pro Leu Ile Gly Phe Ile Gly Arg Leu Glu Glu Gln Lys Gly 400 405 410	
TCA GAT ATT CTT GTT GCT GCA ATT CAC AAG TTC ATC GGA TTC Ser Asp Ile Leu Ala Val Ala Ile His Lys Phe Ile Gly Let 415 420 425	د
GAT GTT CAA ATT GTA GTC CTT GTAAGTACCA AATGGACTCA Asp Val Gln Ile Val Val Leu 430	1875
TGGTATCTOT CTTGTTGAGT TTACTTGTGC CGARACTGAA ATTGACCTGC TACTCATOOT ATGCATCAS GGA ACT GGC AAA AAG GAG TTT GAG Gly Thr Gly Lys Lys Glu Fre Glu 435 440	1925 1968

CAG GAG ATT GAA CAG CTC GAA GTG TTG TAC CCT AAC AAA GCT Gln Glu Ile Glu Gln Leu Glu Val Leu Tyr Pro Asn Lys Ala 445 450	2010
AAA GGA GTG GCA AAA TTC AAT GTC CCT TTG GCT CAC ATG ATC Lys Gly Val Ala Lys Phe Asn Val Pro Leu Ala His Met Ile 455	2052
ACT GCT GGT GCT GAT TTT ATG TTG GTT CCA AGC AGA TTT GAA Thr Ala Gly Ala Asp Phe Met Leu Val Pro Ser Arg Phe Glu 470 475 480	2094
CCT TGT GGT CTC ATT CAG TTA CAT GCT ATG CGA TAT GGA ACA Pro Cys Gly Leu Ile Gln Leu His Ala Met Arg Tyr Gly Thr 485 490 495	2136
GTAAGAACCA GAAGAGCTTG TACCTTTTTA CTGAGTTTTT AAAAAAAGAA TCATAAGACC TTGTTTTCCA TCTAAAGTTT AATAACCAAC TAAATGTTAC TGCAGCAAGC TTTTCATTTC TGAAAATTGG TTATCTGATT TTAACGTAAT CACATGTGAG TCAG GTA CCA ATC TGT GCA TCG ACT GGT GGA CTT Val Pro Ile Cys Ala Ser Thr Gly Gly Leu 500 505	2186 2236 2286 2330
GTT GAC ACT GTG AAA GAA GGC TAT ACT GGA TTC CAT ATG GGA Val Asp Thr Val Lys Glu Gly Tyr Thr Gly Phe His Met Gly 510 515 520	2372
GCC TTC AAT GTT GAA GTATGTGATT TTACATCAAT TGTGTACTTG Ala Phe Asn Val Glu 525	2417
TACATGGICO ATTOTOGICT TGATATACCO CTIGITGCAT AAACATTAAC TTATTGCIID TTGAATTIGG TIAG TGC GAT GIT GIT GAC CCA GCT Cys Asp Val Val Asp Pro Ala 530	2467 2512
GAT GTG CTT AAG ATA GTA ACA ACA GTT GCT AGA GCT C Asp Val Leu Lys lle Val Thr Thr Val Ala Arg Ala 535 540	2549

26

SEQ ID No. 3

Sequenced molecule: genomic DNA

Name: GBSS gene fragment from potato

Length of sequence: 492 bp

GAG CTC TCC Glu Leu Ser 565		AGTGTGA ATTTGATAAT	TTGCGTAGGT	45
ACTTCAGTTT G TGCAG	GAA CCT G	GCACTGA TGGATTCCAA CC AAG AAA TGG GAG .a Lys Lys Trp Glu 575	ACA TTG	95 127
CTA TTG GGC Leu Leu Gly 580	TTA GGA GCT To Leu Gly Ala S	CT GGC AGT GAA CCC er Gly Ser Glu Pro 585	GGT GTT GAA Gly Val Glu 590	169
Gly Glu Glu		TT GCC AAG GAA AAT eu Ala Lys Glu Asn 600		211
CCT TAA P Pro *** 606	TGAGCTTTG GTT	ATCCTTG TTTCAACAAT	AAGATCATTA	257
ATCATCTACA A TGTAAAATCC T TGGATCAAAG T	LAATGATTGG TTT TGGTTAATGT TTT	TATGTAG AACCCTATTA TTGCTGG GGAGCAGCAG TGTAGGT AAGGGCTATT GTTATTA CTAACGTTTG ACTAGAA CTAGT	CATATAAGGC TAAGGTGGTG	307 357 407 457 492

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27

SEQ ID No. 4

Sequenced molecule: genomic DNA

Name: Promoter for th GBSS gene from potato

Length of sequence: 987 bp

AAGCTTTAAC	GAGATAGAAA	ATTATGTTAC	TCCGTTTTGT	TCATTACTTA	50
ACAAATGCAA	CAGTATCTTG	TACCAAATCC	TTTCTCTCTT	TTCAAACTTT	100
TCTATTTGGC	TGTTGACGGA	GTAATCAGGA	TACAAACCAC	AAGTATTTAA	150
TTGACTCCTC	CGCCAGATAT	TATGATTTAT	GAATCCTCGA	AAAGCCTATC	200
CATTAAGTCC	TCATCTATGG	ATATACTTGA	CAGTATCTTC	CTGTTTGGGT	250
ATTTTTTTT	CCTGCCAAGT	GGAACGGAGA	CATGTTATGA	TGTATACGGG	300
AAGCTCGTTA	ALAALAATA	CAATAGGAAG	AAATGTAACA	AACATTGAAT	350
GTTGTTTTTA	ACCATCCTTC	CTTTAGCAGT	GTATCAATTT	TGTAATAGAA	400
CCATGCATCT	CAATCTTAAT	LCTAAAATGC	AACTTAATAT	AGGCTAAACC	450
CCATGCATCT	AATGTATTCA	ACCTTTAGAA	TTGTGCATTC	ATAATTAGAT	500
CTTGTTTGTC	CTAALLATT	LCARATATA	TTTACAGTAA	TTTGGAATAC	550
AAAGCTAAGG	GGGAAGTAAC	TAATATTCTA	GTGGAGGGAG	GGACCAGTAC	600
	ATATTATTTT	TAATTACTAT	TTAATAATA	TAATTAACAC	650
CAGTACCIAG		CGTACCGTAG	GAGGGAGTTG	GTTTAGTTTT	700
GAGACATAGG	AATGTCAAGT		CATTGCAAGG	CCAAGTTGAA	750
TTAGATACTA	GGAGACAGAA	CCGGACGGCC	•••••	TCGATGAGCA	800
GTCCAGCCGT	GAATCAACAA	AGAGAGGGCC	CATAATACTG		850
TTTCCCTATA	ATACAGTGTC	CACAGTTGCC	TTCTGCTAAG	GGATAGCCAC	
CCGCTATTCT	CTTGACACGT	GTCACTGAAA	CCTGCTACAA	ATAAGGCAGG	900
CACCTCCTCA	TTCTCACTCA	CTCACTCACA	CAGCTCAACA	AGTGGTAACT	950
TTTACTCATC	TCCTCCAATT	ATTTCTGATT	TCATGCA		987

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28

SEQ ID No. 5

Sequenced molecule: genomic DNA

Name: GBSS gene from potato Length of sequence: 4964 bp

AAGCTTTAAC GAGATAGAAA ATTATGTTAC TCCGTTTTGT TCATTACTTA	50
ACAAATGCAA CAGTATCTIG TACCAAATCC TTTCTCTCTT TTCAAACTTT	100
TCTATTTGGC TGTTGACGGA GTAATCAGGA TACAAACCAC AAGTATTTAA	150
TTGACTCCTC CGCCAGATAT TATGATTTAT GAATCCTCGA AAAGCCTATC	200
CATTAAGTCC TCATCTATGG ATATACTTGA CAGTATCTTC CTGTTTGGGT	250
ATTTTTTTT CCTGCCAAGT GGAACGGAGA CATGTTATGA TGTATACGGG	300
AAGCTCGTTA AAAAAAATA CAATAGGAAG AAATGTAACA AACATTGAAT	350
GTTGTTTTTA ACCATCCTTC CTTTAGCAGT GTATCAATTT TGTAATAGAA	400
CCATGCATCT CAATCTTAAT ACTAAAATGC AACTTAATAT AGGCTAAACC	450
AAGATAAAGT AATGTATTCA ACCTTTAGAA TTGTGCATTC ATAATTAGAT	500
CTTGTTTGTC GTAAAAATT AGAAAATATA TTTACAGTAA TTTGGAATAC	550
AAAGCTAAGG GGGAAGTAAC TAATATTCTA GTGGAGGGAG GGACCAGTAC	600
CAGTACCTAG ATATTATTTT TAATTACTAT AATAATAATT TAATTAACAC	650
GAGACATAGG AATGTCAAGT GGTAGCGTAG GAGGGAGTTG GTTTAGTTTT	760
TTAGATACTA GGAGACAGAA CCGGACGGCC CATTGCAAGG CCAAGTTGAA	750
GTCCAGCCGT GAATCAACAA AGAGAGGGCC CATAATACTG TCGATGAGCA	800
TTTCCCTATA ATACAGTGTC CACAGTTGCC TTCTGCTAAG GGATAGCCAC	850
CCGCTATTCT CTTGACACGT GTCACTGAAA CCTGCTACAA ATAAGGCAGG	900
CACCTCCTCA TTCTCACTCA CTCACTCACA CAGCTCAACA AGTGGTAACT	950
TTTACTCATC TCCTCCAATT ATTTCTGATT TCATGCATGT TTCCCTACAT	1000
TCTATTATGA ATCGTGTTGT GGTGTATAAA CGTTGTTTCA TATCTCATCT	1050
CATCTATTCT GATTTTGATT CTCTTGCCTA CTGTAATCGG TGATAAATGT	1100
GAATGCTTCC TTTCTTCTCA GAAATCAATT TCTGTTTTGT TTTTGTTCAT	1150
CTGTAGCTTA TTCTCTGGTA GATTCCCCTT TTTGTAGACC ACACATCAC	1199
	1241
**** **** **** **** ****	1241
Met Ala Ser Ile Thr Ala Ser His His Phe Val Ser Arg Ser	
1 5 10	
	1202
CAA ACT TCA CTA GAC ACC AAA TCA ACC TTG TCA CAG ATA GGA	1283
Gln Thr Ser Leu Asp Thr Lys Ser Thr Leu Ser Gln Ile Gly	
15 20 25	
CTC AGG AAC CAT ACT CTG ACT CAC AAT GGT TTA AGG GCT GTT	1325
Leu Arg Asn His Thr Leu Thr His Asn Gly Leu Arg Ala Val	
30 35 40	
AAC AAG CIT GAT GGG CTC CAA TCA ACA ACT AAT ACT AAG GTA	1367
Asn Lys Leu Asp Gly Leu Gln Ser Thr Thr Asn Thr Lys Val	
45 50 55	
ACA CCC AAG ATG GCA TCC AGA ACT GAG ACC AAG AGA CCT GGA	
Thr Pro Lys Met Ala Ser Arg Thr Glu Thr Lys Arg Pro Gly	
60 65 70	
TGC TCA GCT ACC ATT GTT TGT GGA AAG GGA ATG AAC TTG ATC	1451
Cys Ser Ala Thr Ile Val Cys Gly Lys Gly Met Asn Leu Ile	
75 80	
TIT GIG GGT ACT GAG GIT GGT CCT IGG AGC AAA ACT GGT GGA	1493
Phe Val Gly Thr Glu Val Gly Pro Trp Ser Lys Thr Gly Gly	_
85 90 95	

CTA GGT GAT GTT CTT GGT GGA CTA CCA CCA GCC CTT GCA Leu Gly Asp Val Leu Gly Gly Leu Pro Pro Ala Leu Ala 100 105 110	1532
GTAAGTCTTT CTTTCATTTG GTTACCTACT CATTCATTAC TTATTTTGTT TAGTTAGTTT CTACTGCATC AGTCTTTTTA TCATTTAG GCC CGC GGA Ala Arg Gly	1582 1629
CAT CGG GTA ATG ACA ATA TCC CCC CGT TAT GAC CAA TAC AAA His Arg Val Met Thr Ile Ser Pro Arg Tyr Asp Gln Tyr Lys 115 120 125	1671
GAT GCT TGG GAT ACT GGC GTT GCG GTT GAG GTACATCTTC Asp Ala Trp Asp Thr Gly Val Ala Val Glu 130	1711
CTATATTGAT ACGGTACAAT ATTGTTCTCT TACATTTCCT GATTCAAGAA TGTGATCATC TGCAG GTC AAA GTT GGA GAC AGC ATT GAA ATT GTT Val Lys Val Gly Asp Ser Ile Glu Ile Val 140 145	1761 1806
CGT TTC TTT CAC TGC TAT AAA CGT GGG GTT GAT CGT GTT TTT Arg Phe Phe His Cys Tyr Lys Arg Gly Val Asp Arg Val Phe 150 160	1848
GTT GAC CAC CCA ATG TTC TTG GAG AAA GTAAGCATAT Val Asp His Pro Met Phe Leu Glu Lys 165 170	1885
TATGATTATG AATCCGTCCT GAGGGATACG CAGAACAGGT CATTTTGAGT ATCTTTTAAC TCTACTGGTG CTTTTACTCT TTTAAG GTT TGG GGC AAA Val Trp Gly Lys 175	1935 1983
ACT GGT TCA AAA ATC TAT GGC CCC AAA GCT GGA CTA GAT TAT Thr Gly Ser Lys Ile Tyr Gly Pro Lys Ala Gly Leu Asp Tyr 180	2025
CTG GAC AAT GAA CTT AGG TTC AGC TTG TTG TGT CAA Leu Asp Asn Glu Leu Arg Phe Ser Leu Leu Cys Gln 190 195 200	2061
GTAAGTTAGT TACTCTTGAT TTTTATGTGG CATTTTACTC TTTTGTCTTT AATCGTTTTT TTAACCTTGT TTTCTCAG GCA GCC CTA GAG GCA CCT Ala Ala Leu Glu Ala Pro 205	2111 2157
AAA GTT TTG AAT TTG AAC AGT AGC AAC TAC TTC TCA GGA CCA Lys Val Leu Asn Leu Asn Ser Ser Asn Tyr Phe Ser Gly Pro 210 215 220	2199

TAT G GTAATTAACA CATCCTAGTT TCAGAAAACT CCTTACTATA Tyr G	2243
TCATTGTAGG TAATCATCTT TATTTTGCCT ATTCCTGCAG GA GAG GAT ly Glu Asp 225	2291
GTT CTC TTC ATT GCC AAT GAT TGG CAC ACA GCT CTC ATT CCT Val Leu Phe Ile Ala Asn Asp Trp His Thr Ala Leu Ile Pro 230 235	2333
TGC TAC TTG AAG TCA ATG TAC CAG TCC AGA GGA ATC TAC TTG Cys Tyr Leu Lys Ser Met Tyr Gln Ser Arg Gly Ile Tyr Leu 240 245 250	2375
AAT GCC AAG GTAAAATTTC TTTGTATTCA CTCGATTGCA Asn Ala Lys 255	2414
CGTTACCCTG CAAATCAGTA AGGTTGTATT AATATATGAT AAATTTCACA TTGCCTCCAG GTT GCT TTC TGC ATC CAT AAC ATT GCC TAC CAA Val Ala Phe Cys Ile His Asn Ile Ala Tyr Gln 260 265	2464 2507
GGT CGA TIT TCT TTC TCT GAC TTC CCT CTT CTC AAT CTT CCT Gly Arg Phe Ser Phe Ser Asp Phe Pro Leu Leu Asn Leu Pro 270 275 280	2549
GAT GAA TTC AGG GGT TCT TTT GAT TTC ATT GAT GGG TAT Asp Glu Phe Arg Gly Ser Phe Asp Phe Ile Asp Gly Tyr 285 290	2588
GTATTTATEC TTGAAATCAG ACCTCCAACT TTTGAAGCTC TTTTGATGCT AGTAAATTGA GTTTTTAAAA TTTTGCAGAT ATGAG AAG CCT GTT AAG Lys Pro Val Lys 295	2638 2685
GGT AGG AAA ATC AAC TGG ATG AAG GCT GGG ATA TTA GAA TCA Gly Arg Lys Ile Asn Trp Met Lys Ala Gly Ile Leu Glu Ser 300 305 310	2727
CAT AGG GTG GTT ACA GTG AGC CCA TAC TAT GCC CAA GAA CTT His Arg Val Val Thr Val Ser Pro Tyr Tyr Ala Gln Glu Leu 315 320 325	2769
GTC TCT GCT GTT GAC AAG GGA GTT GAA TTG GAC AGT GTC CTT Val Ser Ala Val Asp Lys Gly Val Glu Leu Asp Ser Val Leu 330	2811
CGT AAG ACT TGC ATA ACT GGG ATT GTG AAT GGC ATG GAT ACA Arg lys Thr Cys Ile Thr Gly Ile Val Ash Gly Met Asp Thr 345	2853

CAA GAG TGG AAC CCA GCG ACT GAC AAA TAC ACA GAT GTC AAA Gln Glu Trp Asn Pro Ala Thr Asp Lys Tyr Thr Asp Val Lys 355 360 365	2895
TAC GAT ATA ACC ACT GTAAGATAAG ATTTTTCCGA CTCCAGTATA Tyr Asp lle Thr Thr 370	2940
TACTAAATTA TTTTGTATGT TTATGAAATT AAAGAGTTCT TGCTAATCAA AATCTCTATA CAG GTC ATG GAC GCA AAA CCT TTA CTA AAG GAG Val Met Asp Ala Lys Pro Leu Leu Lys Glu 375 380	2990 3033
GCT CTT CAA GCA GCA GTT GGC TTG CCT GTT GAC AAG AAG ATC Ala Leu Gln Ala Ala Val Gly Leu Pro Val Asp Lys Lys Ile 385	
CCT TTG ATT GGC TTC ATC GGC AGA CTT GAG GAG CAG AAA GGT Pro Leu Ile Gly Phe Ile Gly Arg Leu Glu Glu Gln Lys Gly 400 405	
TCA GAT ATT CTT GTT GCT GCA ATT CAC AAG TTC ATC GGA TTG Ser Asp Ile Leu Ala Val Ala Ile His Lys Phe Ile Gly Leu 415 420 425	
GAT GTT CAA ATT GTA GTC CTT GTAAGTACCA AATGGACTCA Asp Val Gln Ile Val Val Leu 430	3200
TGGTATCTCT CTTGTTGAGT TTACTTGTGC CGAAACTGAA ATTGACCTGC TACTCATCCT ATGCATCAG GGA ACT GGC AAA AAG GAG TTT GAG Gly Thr Gly Lys Lys Glu Phe Glu 435 440	3250 3293
CAG GAG ATT GAA CAG CTC GAA GTG TTG TAC CCT AAC AAA GCT Gln Glu Ile Glu Gln Leu Glu Val Leu Tyr Pro Asn Lys Ala 445	
AAA GGA GTG GCA AAA TTC AAT GTC CCT TTG GCT CAC ATG ATC Lys Gly Val Ala Lys Phe Asn Val Pro Leu Ala His Met Ile 455 460 465	
ACT GCT GGT GCT GAT TTT ATG TTG GTT CCA AGC AGA TTT GAA Thr Ala Gly Ala Asp Phe Met Leu Val Pro Ser Arg Phe Glu 470 475 480	
CCT TGT GGT CTC ATT CAG TTA CAT GCT ATG CGA TAT GGA ACA Pro Cys Gly Leu Ile Gln Leu His Ala Met Arg Tyr Gly Thr 485 490 495	
GTAAGAACCA GAAGAGCTTG TACCTTTTTA CTGAGTTTTT AAAAAAAGAA TCATAAGACC TTGTTTTCCA TCTAAAGTTT AATAACCAAC TAAATGTTAC TGCAGCAAGC TTTTCATTTC TGAAAATTGG TTATCTGATT TTAACGTAAT	3511 3561 3611

		32	
CACATGTGAG TCAG		TCG ACT GGT GGA CTT Ser Thr Gly Gly Leu 505	3655
		GGA TTC CAT ATG GGA Gly Phe His Met Gly 520	3697
GCC TTC AAT GTT Ala Phe Asn Val	GAA GTATGTGATT TT Glu 525	ACATCAAT TGTGTACTTG	3742
	ATTTGG TTAG TGC GAT	TTGTTGCAT AAACATTAAC GTT GTT GAC CCA GCT Val Val Asp Pro Ala 530	3792 3837
		GCT AGA GCT CTT GCA Ala Arg Ala Leu Ala 545	3879
		ATG ATA AAA AAT TGC Met Ile Lys Asn Cys 560	3921
	CTC TCC TGG AAG GI Leu Ser Trp Lys 565	FAAGTGIGA ATTTGATAAT	3965
	G GAA CCT GCC AAG	CAGCACTGA TGGATTCCAA S AAA TGG GAG ACA TTG Lys Trp Glu Thr Leu 575	4015 4057
		GAA CCC GGT GTT GAA r Glu Pro Gly Val Glu 590	4099
	Ala Pro Leu Ala Lys	G GAA AAT GTA GCC ACT S Glu Asn Val Ala Thr 605	4141
CCT TAA ATGA Pro *** 606	GCTTTG GTTATCCTTG T	ITCAACAAT AAGATCATTA	4187
ATCATCTACA AAAT TGTAAAATCC TGGT TGGATCAAAG TCAA CTTAGTAATG TAGC GTGAATTTGT TGTA TGAAAATTAC CCAA	GATTGG TTTTTGCTGG GG TAATGT TTTTGTAGGT AL TAGAAA ATAGTTATTA CT ATAAAAT AATAGTAATA TAAAAT AATAGTAATA TAAAAT AATAGTAATA TAAAAT AATAGTTTGTTGGAAAAT GGAAAT AATGTTTGTTGGAAAT	ACCCTATTA TGGGGTCTCA GAGCAGCAG CATATAAGGC AGGGCTATT TAAGGTGGTG TAACGTTTG CAACTAAATA TAGTAGCTA ATATATATGC ATTTGCAGT ACATATATAA GTCGTCCT CCTTTGATGA GTTATAAAT	4287 4337

AGGCTAACTT TGAGGAGATG GCTATTGAAT TTCAAAATGA TTATGTGAAA	4687
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TGTCTCTTGT CTACTACTCT ATTGGTAGTA GTAGTAGTAA TTTTACAATC	4887
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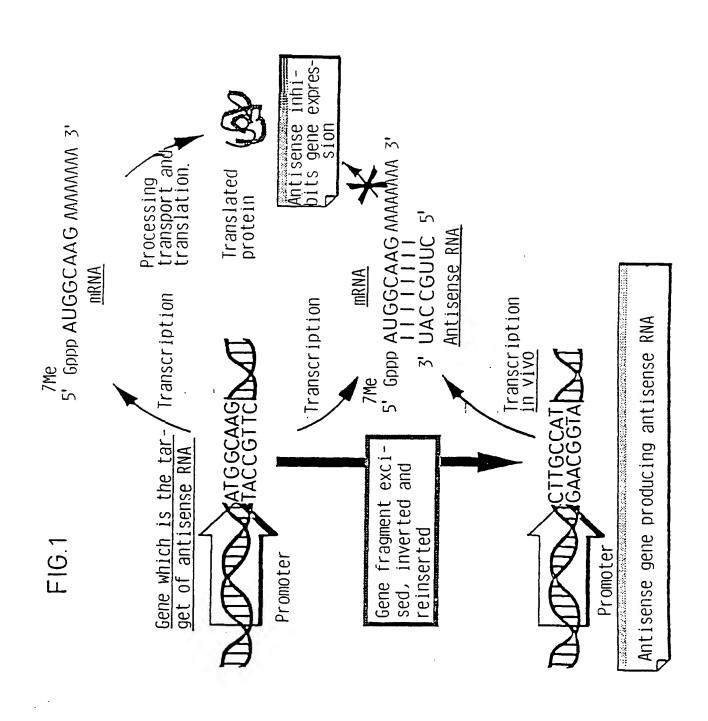
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CLAIMS

- 1. Method of suppressing amylose formation in 5 potato, c h a r a c t e r i s e d by genetically engineered modification of the potato by introducing into the genome of the potato tissue a gene construct comprising a fragment of the potato gene which codes for formation of granule-bound starch synthase (GBSS gene) inserted in the 10 antisense direction, said fragment being selected among the fragments which essentially have the nucleotide sequences stated in SEQ ID No. 1, SEQ ID No. 2 and SEQ ID No. 3 together with a promoter selected among CaMV 35S, patatin I and the GBSS promoter.
- 2. Amylopectin-type native starch, c h a r a c t e r i s e d in that it has been obtained from potato which has been modified in a genetically engineered manner for suppressing formation of amylose-type starch.
- 3. Derivatised amylopectin-type starch, c h a r -20 a c t e r i s e d in that it is amylopectin-type starch extracted from potato which has been modified in a genetically engineered manner for suppressing formation of amylose-type starch, said amylopectin-type starch subsequently being derivatised in a chemical, physical or enzymatic manner.
- 4. Fragment of the gene coding for granule-bound starch synthase (GBSS) in potato, said fragment being selected among the fragments which essentially have the nucleotide sequences stated in SEQ ID No. 1, SEQ ID No. 2 30 and SEQ ID No. 3.
 - 5. Promoter for the gene for granule-bound starch synthase (GBSS) in potato, said promoter being tuber-specific and having essentially the nucleotide sequence stated in SEQ ID No. 4.
- 6. Gene coding for granule-bound starch synthase in potato (GBSS gene) having essentially the nucl otide sequence stated in SEQ ID No. 5.

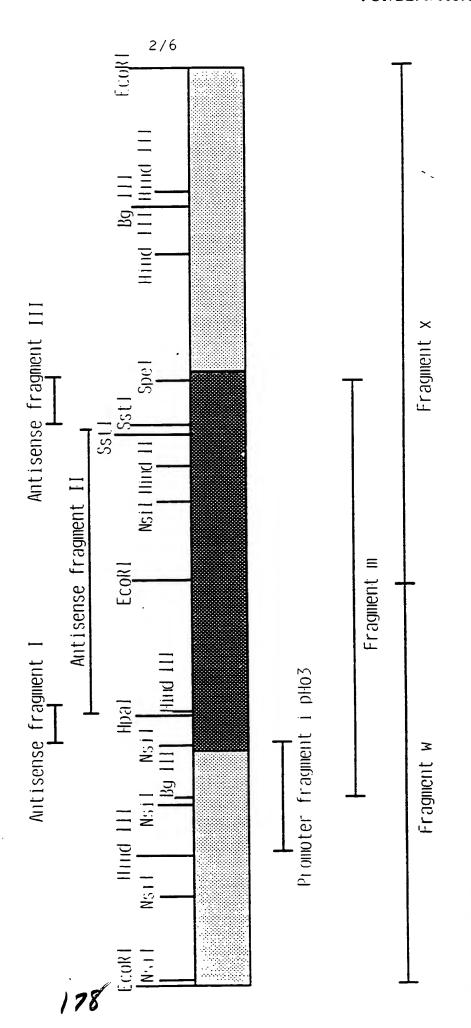
- 7. Antisense construct for inhibiting expression of the gene for granule-bound starch synthase in potato, comprising
- a) a promoter,
- 5 b) a fragment of the gene coding for granule-bound starch synthase inserted in the antisense direction, said fragment being selected among the fragments having essentially the nucleotide sequences stated in SEQ ID No. 1, SEQ ID No. 2 and SEQ ID No. 3.
- 8. Antisense construct as claimed in claim 7, c h a r a c t e r i s e d in that the promoter essentially has the sequence stated in SEQ ID No. 4.
- 9. Antisense construct as claimed in claim 7, c h a r a c t e r i s e d in that the promoter is select15 ed among the CaMV 35S promoter and the patatin I promoter.
 - 10. Vector comprising a fragment of the gene coding for granule-bound starch synthase (GBSS) in potato, said fragment being selected among the fragments having essentially the nucleotide sequences stated in SEQ ID No. 1,
- 20 SEQ ID No. 2 and SEQ ID No. 3, and inserted in the antisense direction.
 - 11. Vector comprising the antisense construct as claimed in any one of claims 7-9.
- 12. Cell of potato plant whose genome comprises the 25 antisense construct as claimed in any one of claims 7-9.
 - 13. Potato plant whose genome comprises the antisense construct as claimed in any one of claims 7-9.
 - 14. Potato tubers whose genome comprises the antisense construct as claimed in any one of claims 7-9.
- 15. Seeds from potato plant, whose genome comprises the antisense construct as claimed in any one of claims 7-9.
 - 16. Microtubers of potato, whose genome comprises the antisense construct as claimed in any one of claims 7-9.

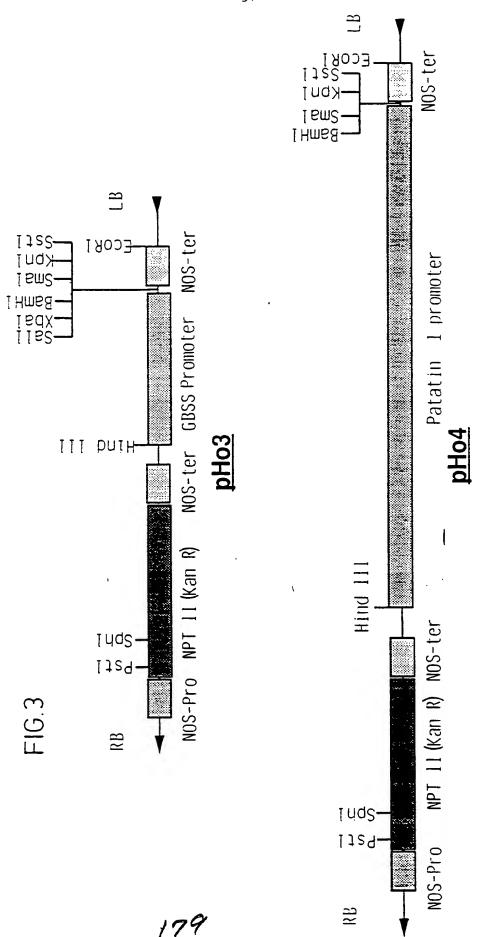
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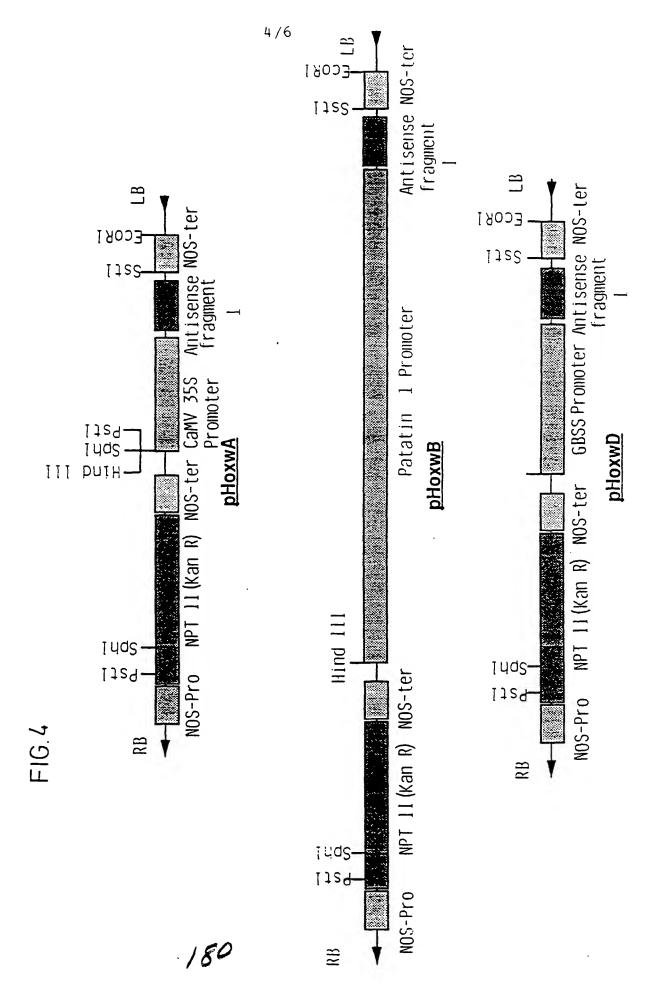


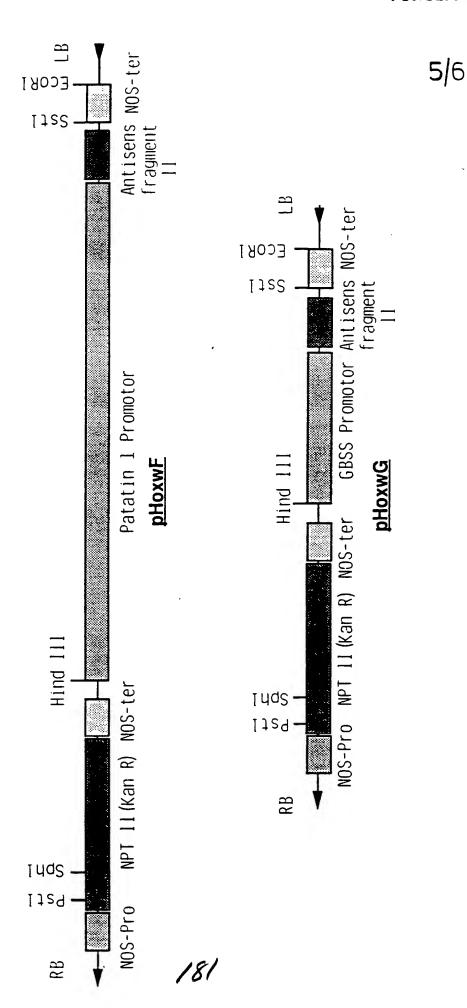
1.0 kb

Result of restriction analysis. GBSS coding region including introns are marked in a darker tone.



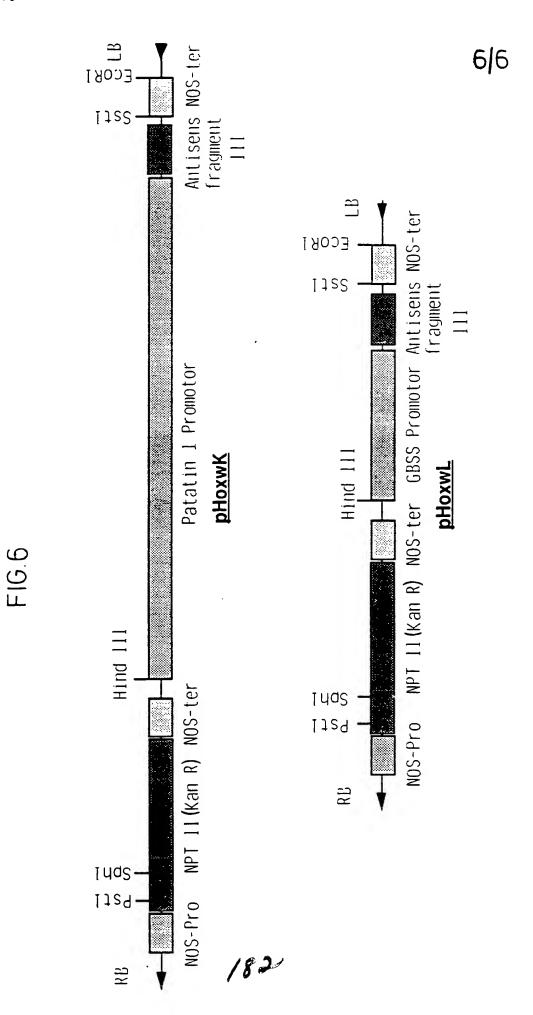






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INTERNATIONAL SEARCH REPORT

International Application No. PC

PCT/SE 91/00892

I. CLASS	IFICATION OF SUBJECT MATTER (if several classifi		3E 91/00892		
According	to International Patent Classification (IPC) or to both No				
IPC5: C	12 N 15/56, 9/42, A 01 H 5/00				
II FIFT DS	SEARCHED				
11. 712250	Minimum Documer	ntation Searched?			
Classification	on System C	lassification Symbols			
IPC5	C 12 N; A 01 H				
	Documentation Searched other	than Minimum Documentation			
	to the Extent that such Documents	are included in Fields Searched ⁸			
SE,DK,F	I,NO classes as above				
	MENTS CONSIDERED TO BE RELEVANT9				
Category *	Citation of Document, ¹¹ with indication, where app	rongiste of the selevent page 22	Belowet to Claim to 13		
			Relevant to Claim No. ¹³		
P,X	MOL GEN GENET, Vol. 225, 1991 R. al: "Inhibition of the expre		1-16		
	for granule-bound starch syr				
antisense constructs", see page 289 -					
	page 296	. 5			
	ED 40 0000500 (IMPERIAL CUENT)	AL INDUCTOIS DIO			
A	EP, A2, 0368506 (IMPERIAL CHEMIC 16 May 1990.	CAL INDUSTRIES PLC)	1-16		
Ì	see especially claim 14				
Α	PLANT SCIENCE, Vol. 64, 1989 R.O	3.F. Visser et	1-16		
	al: "Molecular cloning and p				
	characterization of the gene				
	starch synthase from a wildt				
	<pre>amylose-free potato(solanum cited in the application</pre>	tuberosumi.)",			
İ	<i>,</i>				
	categories of cited documents: 10	"To later document published after or priority date and not in confl	the international filing date		
A docu	"A" document defining the general state of the art which is not considered to be of particular relevance or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention				
"E" earli filin	er document but published on or after the international g date	"X" document of particular relevance	e, the claimed invention		
"L" dock	ment which may throw doubts on priority claim(s) or this cited to establish the publication date of another	involve an inventive step	annot be considered to		
citat	ion or other special reason (as specified)	"Y" document of particular relevance cannot be considered to involve	an inventive step when the		
"O" docu	iment referring to an oral disclosure, use, exhibition or r means	document is combined with one ments, such combination being in the art.	or more other such docu- obvious to a person skilled		
P docu	ment published prior to the international filing date but than the priority date claimed		patent family		
IV. CERTIF	TCATION				
Date of the	Actual Completion of the International Search	Date of Mailing of this International S	earch Report		
30th Ma	rch 1992	1992 -04- 0 1			
Internationa	l Searching Authority	Signature of Authorized Officer			
		Udiail & Berry	フ		
	SWEDISH PATENT OFFICE	Mikael G:son Bergstra	ınd		
Form PCT/IS/	J/210 (second sheet) (January 1985)				

International Application No. PCT/SE 91/00892

International Application No. PCI/SE 91/00092				
III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FR M THE SECOND SHEET) Category Citation of Document, with indication, where appropriate, of the relevant passages Relevant to Citation of Document, with indication, where appropriate, of the relevant passages				
Category	Citation of Document, with more appropriate, or the relevant passages	Relevant to Claim No		
A	EP, A2, 0335451 (VERENIGING VOOR CHRISTELIJK WETENSCHAPPELIJK ONDERWIJS) 4 October 1989, see the whole document	1-16		
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ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO.PCT/SE 91/00892

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the Swedish Patent Office EDP file on 28/02/92. The Swedish Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

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Patent document cited in search report	Publication date	Patent family member(s)		Publication date
EP-A2- 0368506		AU-D- 4430789 JP-A- 2273177		90-08-16 90-11-07
EP-A2- 0335451	89-10-04	JP-A- NL-A-	2016985 8800756	90-01-19 89-10-16



IDENTIFICATION OF INTERNATIONAL APPLICATION	Applicant's or Agent's File Reference
DENTIFICATION OF INTERNATIONAL APPLICATION	2912327
International Application No.	International Filing Date
PCT/SE 91/00892	20th December 1991
Receiving Office	Priority Date Claimed
RO/SE	21st December 1990
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Applicant AMYLOGENE HB et al	
I. CERTAIN CLAIMS WERE FOUND UNSEARCHABLE 1 (Observations on supplemental sheet (2))
II. UNITY OF INVENTION LACKING 2 (Observations on supp	plemental sheet (2))
III. TITLE, ABSTRACT AND FIGURE OF DRAWING	
1. The following indicated items are approved as submitted by the X Title.	applicant: 3
	and the state of t
2. The texts established by this International Searching Authority Title.	of the following indicated items are set forth below:
Abstract.	
Abstract.	
Text of the abstract continued on supplemental sheet (1)	
	and the second transfer of the second transfe
	this International Searching Authority as proposed in form
 This report is incomplete as far as the abstract is concern prepared by this International Searching Authority has no 	ned as the time limit for comments by the applicant on the draft it expired.4
4. Figure to be published with abstract ⁵	
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△ as suggested by the applicant	_
because the applicant failed to suggest a figure	
herause this figure better characterizes the invention	

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See notes on accompanying sheet

INTERNATIONAL SEARCH REPORT

International Application No PCT/SE 91/00892

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		15/56, 9/42, A 01 H 5/00		
II. FIELD	S SEARCH			
		Minimum Documer		
Classificati	ion System	c	tassification Symbols	
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IPC5		C 12 N; A 01 H		
		Documentation Searched other to the Extent that such Documents		
SE,DK,	FI,NO c	lasses as above		
III. DOCU	MENTS CO	DISIDERED TO BE RELEVANT 9		
Category *		on of Document, ¹¹ with indication, where app	ropriate, of the relevant passages 12	Relevant to Claim No.13
P,X	 	N GENET, Vol. 225, 1991 R.	 	1-16
,^	al fo an	: "Inhibition of the express granule-bound starch syntisense constructs", see tige 296	ession of the gene othase in potato by	1 10
A	16	, 0368506 (IMPERIAL CHEMIC May 1990, ee especially claim 14 	CAL INDUSTRIES PLC)	1-16
A	al ch st	SCIENCE, Vol. 64, 1989 R.G.: "Molecular cloning and paracterization of the generated synthase from a wildinglose-free potato(solanum ted in the application ——	partial e for granule-bound type and an	1-16
"A" doc con "E" ear "L" doc whi cits "O" doc oth "P" doc late	cument definition of the control of the cited course of the cited	es of cited documents: 10 ning the general state of the art which is not be of particular relevance ent but published on or after the international th may throw doubts on priority claim(s) or to establish the publication date of another er special reason (as specified) rring to an oral disclosure, use, exhibition or ished prior to the international filing date but priority date claimed	"T" later document published after or priority date and not in conficited to understand the princip invention "X" document of particular relevant cannot be considered novel or involve an inventive step "Y" document of particular relevant cannot be considered to involve document is combined with one ments, such combination being in the art. "&" document member of the same	ce, the claimed invention cannot be considered to ce, the claimed invention e an invention e e an inventive step when the cor more other such docupovious to a person skilled patent family
	SWED	g Authority ISH PATENT OFFICE	Signature of Authorized Office Willaud Heyr Mikael G:son Bergstr	
Form PCT/IS		and sheet) (January 1985)	See note:	s on accompanying sheet

International Application No. PCT/SE 91/00892

	International Application No. PC1/3E 91/00032				
III. DOCL	MENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET) Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No			
A .	EP, A2, 0335451 (VERENIGING VOOR CHRISTELIJK WETENSCHAPPELIJK ONDERWIJS) 4 October 1989, see the whole document	1-16			
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ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO.PCT/SE 91/00892

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The Swedish Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
P-A2- 0368506		AU-D- JP-A-	4430789 2273177	90-08-16 90-11-07
P-A2- 0335451	89-10-04	JP-A- NL-A-	2016985 8800756	90-01-19 89-10-16